Swine Day 2017 Rowland Lab Research Update

Raymond (Bob) Rowland K-State Swine Day Progress Report November 17, 2016 Manhattan





North American PRRS Symposium on PRRS, Emerging and Foreign Animal Diseases and National Swine Improvement Federation NAPRRS-NSIF Joint Conference

December 1-3, 2017 Intercontinental Chicago Magnificent Mile

November 24, 2017- Conference registration deadline





Genetic approaches for improving swine health in response to PRRSV infection Raymond (Bob) Rowland- Kansas State University browland@vet.k-state.edu

Collaborators

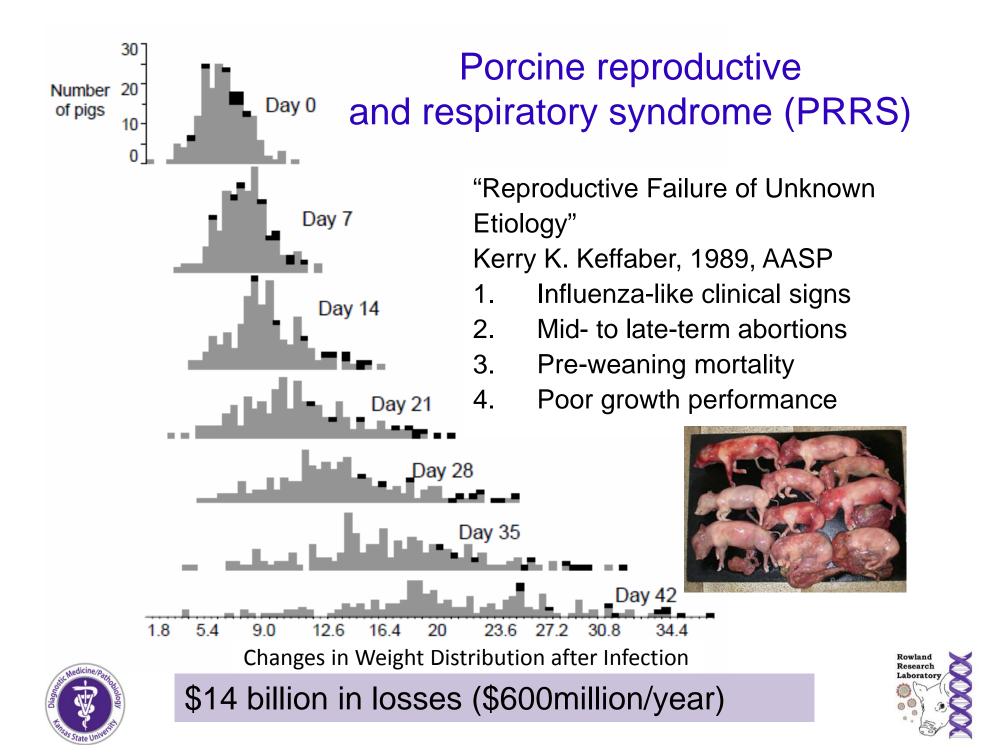
Randy Prather, University of Missouri- Genetically modified pigs that are disease resistant

Jack Dekkers, Iowa State University- Genomic markers for breeding disease resistance

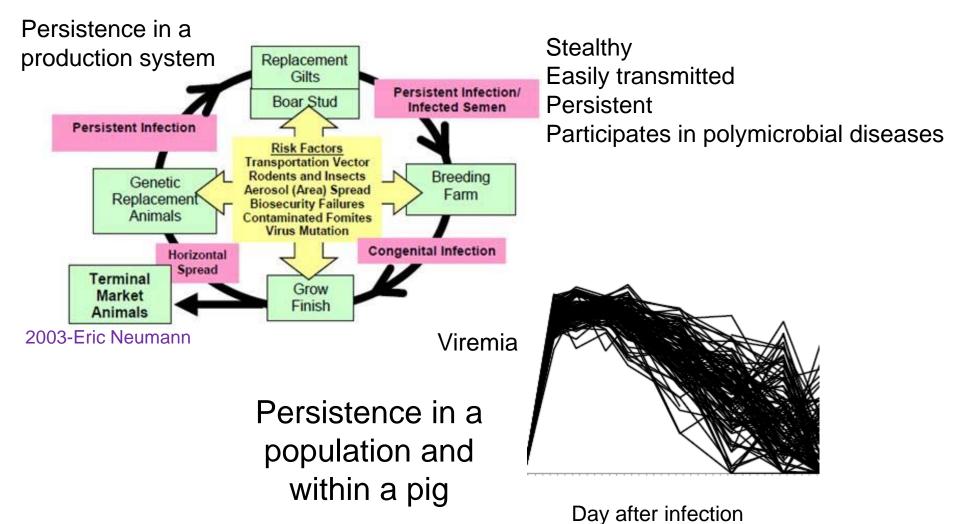
Joan Lunney, ARS-USDA- Genetics of the response of pigs to infection







PRRS is a production system disease Endemic phase with outbreaks of severe disease



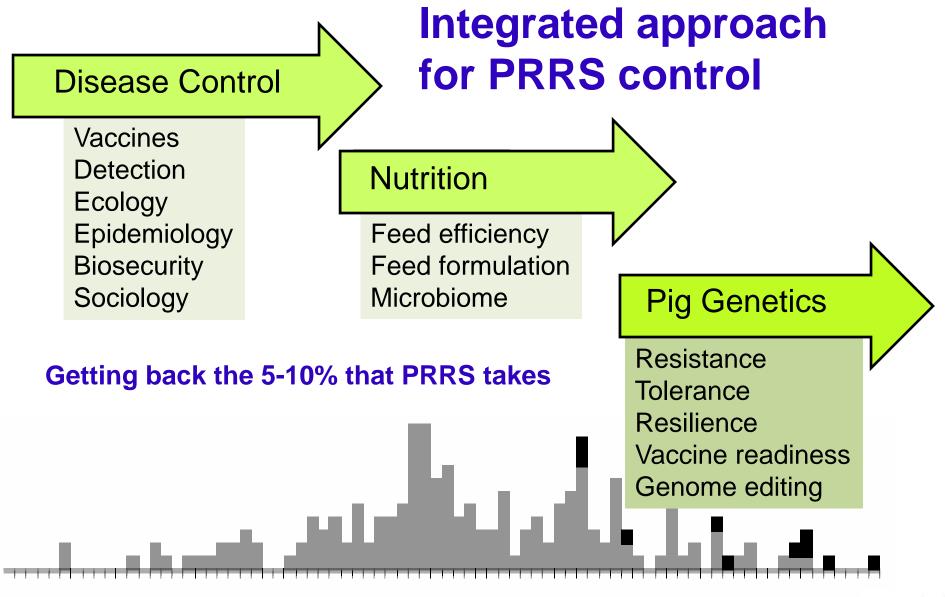
















PRRS vaccines

- Modified live virus (MLV) vaccine introduced in the U.S.1994- approved for use in PRRSV-infected herds
- MLV limitations-virus shedding, persistent infection, incomplete immune protection, inability to differentiate infected from vaccinated animals (DIVA), potential for reversion to virulence
- Killed vaccines are not effective
- Subversion of host immunity and antigenic variation have made further advances in vaccines difficult to achieve

Conclusions: Vaccines are a poor option for disease control and eradication- Vaccinated animals cannot be transported to PRRSV-free regions.





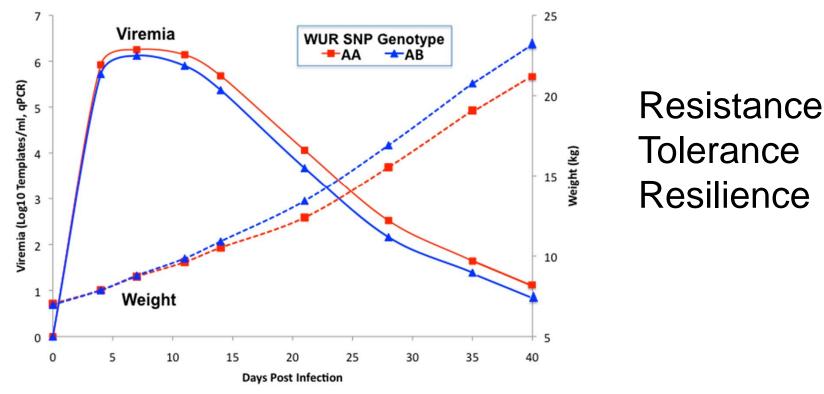
The application of genetics for improving animal health

•Marker selected breeding to improving response

 Modify genes involved in response to infection Insertion of genes to promote resistance
<u>Deletion of genes involved in virus susceptibility</u>

"Genome Editing"

The favorable SSC4 marker, WUR, results in a 10% increase in weight and a decrease in viremia



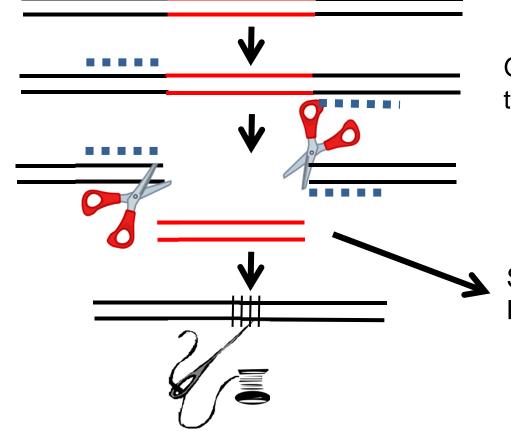
Boddicker, et al. 2012. Evidence for a major QTL associated with host response to porcine reproductive and respiratory syndrome virus challenge. J Anim Sci. 90:1733-1746





Genetic modification CRISPR/Cas 9 system

Gene of interest (CD163)



Guide sequences direct were the genome is cut

Molecular scissors cut out DNA segment

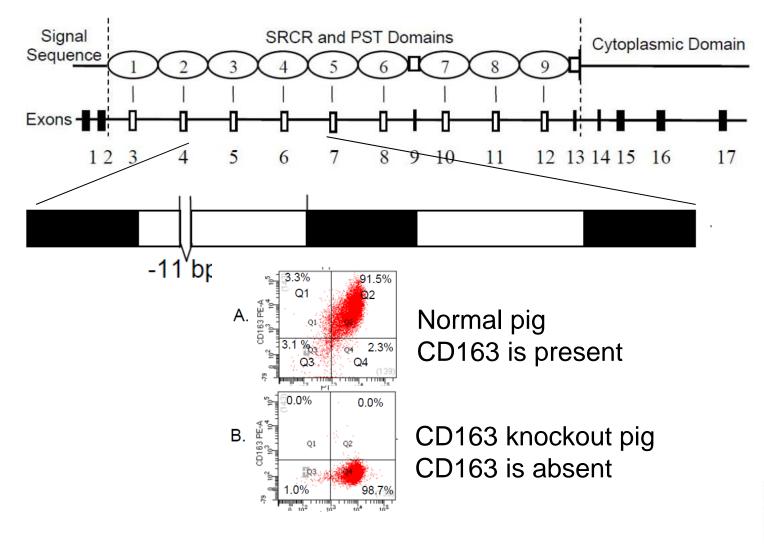
Segment is removed and the DNA ends rejoined







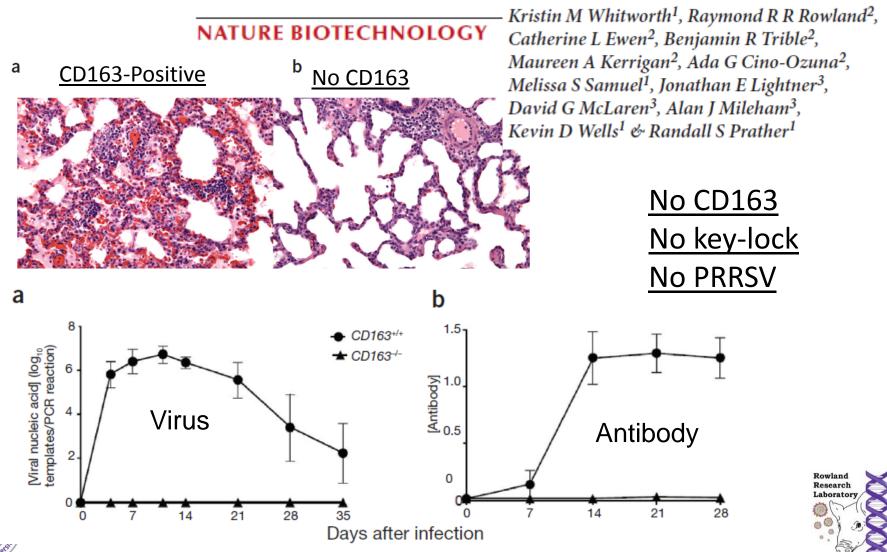
Knocking out CD163 by deleting 11of 2.7 billion bases of the pig genome (Randy Prather)







December 7, 2015 Gene-edited pigs are protected from porcine reproductive and respiratory syndrome virus



Reproductive PRRS Infection of pregnant gilt/sow at 90 days of gestation







Prevention of reproductive PRRS The end of a disease

| Table 1. CD163 parental and fetal genotypes used in this study | | | | | | | | | |
|--|------|----------|-------|-------------|-------------|---------|--|--|--|
| | CD | 163 Geno | type | | | | | | |
| | Pa | rents | | Day of | Day of | No.of | | | |
| Gilt No. | Male | Dam | Fetus | Infection*1 | Gestation*2 | Fetuses | | | |
| 138 | +/+ | +/+ | +/+ | 91 | 106 | 16 | | | |
| 139* ³ | +/+ | +/+ | +/+ | 91 | 106 | 14 | | | |
| 140 | +/+ | +/+ | +/+ | 91 | 106 | 12 | | | |
| 84 | +/+ | -/- | +/- | 89 | 109 | 14 | | | |
| 87 | +/+ | -/- | +/- | 89 | 109 | 17 | | | |
| 122 | +/+ | -/- | +/- | 89 | 109 | 11 | | | |
| 86 | -/- | -/- | -/- | 90 | 109 | 7 | | | |
| 121 | -/- | -/- | -/- | 90 | 109 | 9 | | | |
| *1 Gestation day when dams were infected | | | | | | | | | |
| * ² Gestation day when fetuses were removed | | | | | | | | | |
| * ³ PRRSV-infected dam aborted at 106 days of gestation | | | | | | | | | |





Prevention of reproductive PRRS The end of a disease

| | Dam | Fetus | Dam No. | Genotype Dam Fetus | Right Uterine Horn Left Uterine Horn | |
|--------|----------------|-------|--------------|-----------------------|---|-----------------------------------|
| ++ | | ++ | 138 (3.6) | +/+ +/+ | $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ | |
| | ŦŦ | | 140 (4.1) | +/+ +/+ | $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ | |
| | | | 84 (N) | -/- +/- | $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ | |
| | | +/- | 87 (N) | -/- +/- <u>1</u> N | $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | |
| | | | 122 (N) | -/- +/- | $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ | |
| - | | _/_ | 86 (N) | -//- | $\begin{array}{c ccccccccccccccccccccccccccccccccccc$ | |
| ingno. | edicine/Pathan | | 121 (N) | -//- | $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | Rowland Research Laboratory |



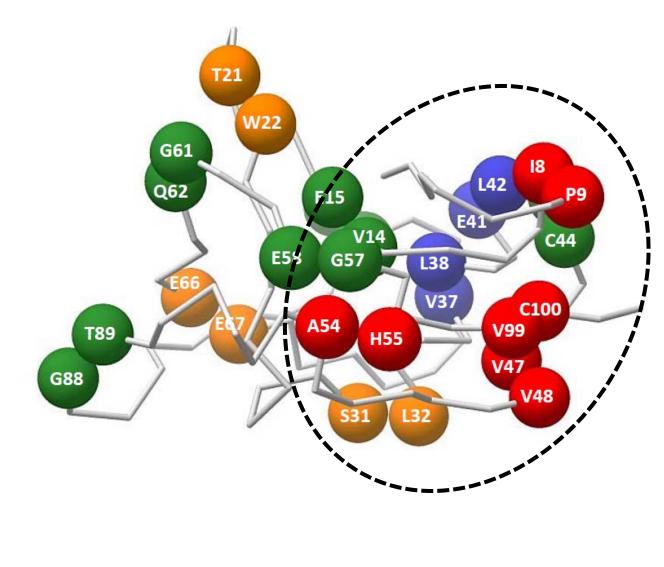
The end of a disease

- The absence of CD163 in the dam surrounds the developing fetus with a protective barrier, preventing infection with PRRSV
- The piglets are born with normal CD163 levels and are susceptible to PRRSV
- Still need that miracle vaccine





Mapping the regions where PRRSV interacts with CD163





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NBAF



The NBAF, a new, state-of-the-art biosafety level (BSL) 3 & 4 facility located in Manhattan, KS, will enable the U.S. to conduct comprehensive research, develop vaccines and anti-virals, and provide enhanced diagnostic capabilities to protect our country from numerous foreign animal, emerging and zoonotic diseases to assist in protecting our food supply and the nation's agriculture economy and public health. NBAF-associated projects in the Rowland lab (Biosecurity Research Institute)

African swine fever virus (ASFV) and classical swine fever virus (CSFV)

Vaccines

Diagnostics

Genetics of disease resistance

Risks for introduction







Co-Directors USDA ARS BARC Joan Lunney Kansas State University RRR (Bob) Rowland Iowa State University Jack Dekkers



Kansas NBAF Transition Fund



- NIFA award #2013-68004-20362
- National Pork Board
- Genome Canada
- LLNL



